

Comparative Assessment of Nutritional, Antioxidant and Phytochemical Properties of Wild Yams (*Dioscorea* spp.) Accessions in Peninsular Malaysia

Mohd Zulkhairi Azid^{1,*}, Mohd Norfaizal Ghazalli¹, Abdul Muhaimin Abdul Kadir¹,
Siti Aisyah Mohd Noor¹, Saidatul Aqilah Mohamad Yusof¹, Nur Daliana Yusof¹,
Mohammad Raiehan Prayoga², Salmaniza Salleh¹

¹Agrobiodiversity and Environment Research Centre, Malaysian Agricultural Research and Development Institute (MARDI),
MARDI Headquarters, Persiaran MARDI-UPM, 43400 Serdang, Selangor, Tel. +603-8953 6325, Fax. +603-8953 6330, Malaysia.

²Agrotechnology Study Program, Faculty of Agriculture, Universitas Muhammadiyah Sumatera Utara, North Sumatra,
Jl. Kap. Mukhtar Basri No. 3 Medan, 20238, North Sumatra, Tel. +61-6619056, Fax. +61-6625474, Indonesia.

Corresponding author*

zulkhairi@mardi.gov.my

Manuscript received: 06 February, 2025. Revision accepted: 20 May, 2025. Published: 16 June, 2025.

Abstract

Yams (*Dioscorea* spp.) are one of the important tuber crops for sustainability and food security in the world. However, *Dioscorea* spp. is considered an underutilized crop and is not heavily consumed, especially by local people in Malaysia, due to its limitations in its nutritional composition and beneficial studies. This study was carried out to evaluate various accessions of wild yams in Peninsular Malaysia. Eight different accessions of yams (*D. alata* (DA1-DA5), *D. alata* var. *purpurea* (DVP), *D. esculenta* (DE) and *D. piscatorum* (DP) were collected from various localities in farmer's farms in Peninsular Malaysia. All the samples were investigated for their proximate composition, polyphenols and flavonoid content and in vitro antioxidant activity. *D. piscatorum* (DP) collected in Kuala Koh, Kelantan, showed the highest amount of carbohydrate content and energy compared to all accessions ($p < 0.05$) with 90.8 g/100g and 374.33 kcal/100g, respectively. Meanwhile, a variety of *D. alata* (DA2) collected in Machang, Kelantan, showed a high amount of protein content (10.57 g/100g) ($p < 0.05$). The flavonoid/phenolic ratio was highest in the methanolic extract of DVP (0.72), followed by DE (0.64) and DP (0.61), respectively. However, the methanolic extract of DP showed the lowest inhibition concentration value ($IC_{50} = 4.73$ mg/ml), indicating its stronger scavenging activity towards DPPH free radicals. The comparison and assessment of different accessions of *Dioscorea* spp. on their nutritional composition and phytochemicals highlights the suitability of each sample to accommodate specific dietary or health-related applications.

Keywords: Antioxidant; Dioscoreaceae; polyphenols; ubi; underutilized.

INTRODUCTION

Dioscorea belongs to the Dioscoreaceae family and is also known as yams, greater yam, "Shanyao" in China, "umbi" or "uwi" in Indonesia, "ube" in the Philippines or "ubi" in Malaysia (Mojiono et al., 2022; Li et al., 2023; Azahana et al., 2023; Ferriol, 2023). The world production share of yams by regions from 2020-2022 in Africa (98%), Americas (1.2%), Oceania (0.5%) and Asia (0.2%), with the top producer countries being Nigeria, Ghana, Côte d'Ivoire and Benin. The increase in yams production worldwide from 81 million tons (2020) to 88 million tons (2022) showed a significant demand for this crop (FAO STAT 2024). Meanwhile, according to the statistics released by the Department of Agriculture of Peninsular Malaysia, the production of *Dioscorea alata* or "ubi badak" in 2023 was estimated at around four metric tons (DOA, 2023). Limited

knowledge of its nutritional value compared to other tuberous crops is one of the reasons this crop is not commercially cultivated and consumed in Malaysia. The *Dioscorea* spp. commonly consumed by local people on the East Coast of Peninsular Malaysia and can be found abundantly in local markets, especially during the monsoon season from October to December each year (Azahana et al., 2023).

The tuber flesh of *Dioscorea* spp. is rich in carbohydrates, making it one of the primary staple foods in certain countries, such as those in Africa, and an alternative starchy food in some areas in Malaysia. The carbohydrate content of several species of *Dioscorea* (*D. alata*, *D. esculenta*, *D. rotunda*, *D. cayenensis* and *D. bulbifera*) was 50% - 77% depending on their varieties and location of cultivation (Oko & Famurewa, 2015; Ezeabara & Anona, 2018; Godfrey et al., 2023; Sato et al., 2024). Meanwhile, the protein content in the various

species of *Dioscorea* was reported in the range of 1.13% - 6.20% (*D. hispida*), 4.7% - 8.99% (*D. alata*), 7.25% (*D. esculenta*) and 8.83% (*D. bulbifera*) (Saleha et al., 2018; N'dri et al., 2018; Yalindua et al., 2021; Rayamajhi et al., 2024). Recent studies on its biological activities from extracts of *Dioscorea* spp. also revealed interesting activities in antioxidant (Adoménienė & Venskutonis, 2022), antimicrobial (Induar et al., 2024), antidiabetic (Rayamajhi et al., 2024), antidiarrheal (Islam et al., 2024), antihypertensive (Logan et al., 2024), hypolipidemic (Li et al., 2019; Povydysh et al., 2023), antiproliferative and anticancer (Mainasara et al., 2021; Nurul Makiyah et al., 2023). The astonishing pharmacological activities of *Dioscorea* spp. mainly due to the presence of its secondary metabolites, such as phenolic compounds, including flavonoids and anthocyanins, steroidal saponin, and carotenoids (Ren et al., 2015; Price et al., 2018; Ukom et al., 2020; Safitri et al., 2021; Kuagny et al., 2023; Syahputra et al., 2024; Islam et al., 2024). However, studies on proximate composition and phytochemicals of various *Dioscorea* spp. accessions in Malaysia remain limited. Thus, this study aims to deliver and assess various accessions of *Dioscorea* spp. collected in Peninsular Malaysia for its

nutritional values, phenolics and flavonoid content, and antioxidant activity.

MATERIALS AND METHODS

Collection and Preparation of the Sample

Eight different accessions of *Dioscorea* spp. were collected across Peninsular Malaysia (Table 1 and Figure 1). Each species was identified by a botanist at the Agrobiodiversity and Environmental Research Centre, MARDI, Serdang. The flesh color and cross-section reviewed under a stereomicroscope are shown in Figure 2. All the specimens were deposited in the Herbarium, Kompleks MyGenebank, MARDI, Serdang. All the samples were cleaned, and the skin was peeled. The samples were cut into small cubes and dried in the industrial oven at 50°C for three days or until the moisture content remained constant. The samples were ground into a fine powder (18 mesh) using a mechanical grinder (IKA Werke MF 10 Basic, Germany). The sample powder was sealed and kept in the chiller at 4°C until further analysis was needed.

Table 1. Various accessions of *Dioscorea* spp. collected in Peninsular Malaysia.

Sample	Sample Code	Location	Tuber Flesh Color
<i>D. alata</i>	DA1	Bukit Payong, Terengganu	Light white yellowish
<i>D. alata</i>	DA2	Machang, Kelantan	Light white yellowish
<i>D. alata</i>	DA3	Pasar Pasir Putih, Kelantan	Light white yellowish
<i>D. alata</i>	DA4	Marang, Terengganu	Light white yellowish
<i>D. alata</i>	DA5	Wakaf Bharu, Kelantan	Light white yellowish
<i>D. alata</i> var. <i>purpurea</i>	DVP	Wakaf Bharu, Kelantan	Dark purple
<i>D. esculenta</i>	DE	Teluk Pelandok, Negeri Sembilan	Light yellow
<i>D. picatorum</i>	DP	Kuala Koh, Kelantan	Dark yellow-Orangish

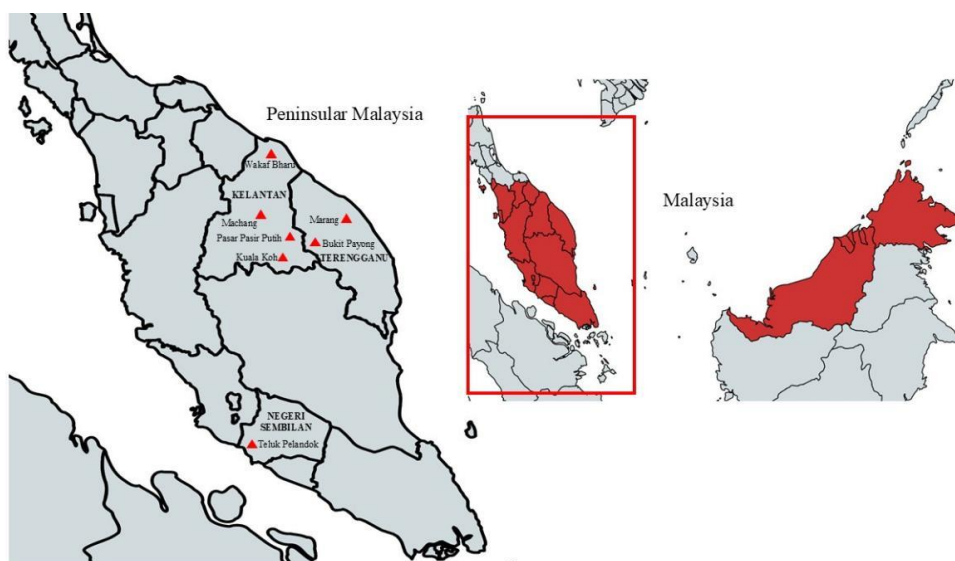


Figure 1. The area of the sample (*Dioscorea* spp.) was collected in Peninsular Malaysia.



Figure 2. Flesh color and structure were reviewed after cross-section under a stereomicroscope of *Dioscorea* spp. collected. a). *Dioscorea alata* (white) b). *Dioscorea alata* var *purpurea* c). *Dioscorea esculenta* and d). *Dioscorea piscatorum*.

Proximate and Nutritional Composition

Approximately 250g of dried samples were weighed and sent to the accredited laboratory for proximate composition analysis. Each reading was done in triplicate.

Total Phenolic Content (TPC)

The total phenolic contents of the sample were determined using the Folin–Ciocalteu method with some modifications (Mirfat et al., 2013). A 2g dried powder sample was extracted in 20 ml of 70% methanol. Crude extract (50µL) was mixed with 100µL of Folin Ciocalteu's phenol reagent (Merck, Germany). After 3 minutes, 100µL of 10% sodium carbonate (Na_2CO_3) (Sigma Aldrich, USA) was added to the reaction mixture and allowed to stand in the dark for 60 minutes. The

absorbance was measured at 725 nm, and the total phenolic contents were calculated from a calibration curve using gallic acid (0-10 µg/mL) as a standard reference. The estimation of phenolic contents was conducted in triplicate. The results were presented as mean values \pm standard deviations and expressed as mg gallic acid equivalent per 100 g of samples (mg GAE/100g) in dry weight (DW).

Total Flavonoid Content (TFC)

The total flavonoid content was measured using the previously published method with minor modifications (Mirfat et al., 2013). Approximately 100µl of extract was diluted with 400 µl of distilled water. Subsequently, a 5% sodium nitrite (NaNO_2) solution (30µl) was added and allowed to react for 5 min. Afterwards, 10%

aluminium chloride ($\text{AlCl}_3 \cdot 6\text{H}_2\text{O}$) (20 μL) was added and left for 5 min. Finally, 200 μL of sodium hydroxide (NaOH) was added, and the mixture was mixed using a vortex. All samples were analyzed in triplicate, and the absorbance was measured immediately at 510 nm. Quercetin was used to calculate the standard curve, and the results were expressed as mg quercetin per 100 g sample (mg QE/100g) in dry weight (DW).

Free Radical Scavenging Assay

All the powder samples underwent extraction and were assessed for their free radical scavenging ability, following the previously described procedure with minor modification (Salahuddin et al., 2020; Zulkhairi et al., 2021). The assay was conducted using a 96-well plate, with 2g of dried powder sample extracted in 20 ml of 70% methanol. The stock solution was diluted to the desired concentration for the working solution. The final volume obtained (7 μL) was mixed with 280 μL of a methanolic solution of 2,2-diphenyl-1-picrylhydrazyl (DPPH) (Sigma, USA). The plate was covered with aluminium foil to prevent exposure to sunlight and was kept in a dark place for 30 minutes. Analysis was performed using a UV spectrophotometer (Eon Biotek

Instrument) at 517 nm. The results were expressed as the inhibition concentration (IC_{50}) value in mg/mL, representing the concentration at which DPPH radicals were scavenged by 50%.

Statistical Analysis

All triplicate samples were analyzed using Duncan's Multiple Range Test (DMRT) (Mean \pm SEM, $n = 3$) with SAS Version 9.4.

RESULTS AND DISCUSSION

Proximate and Nutritional Composition

Proximate and nutritional composition of *Dioscorea* spp. shown in Table 2. Accessions DA1-DA5, DVP and DE showed a range of carbohydrate content <85 g/100g, while DP had significantly the highest carbohydrate content with 90.8 g/100g ($p < 0.05$). Additionally, DP also provides significantly ($p < 0.05$) higher energy in 100g compared to other *Dioscorea* accessions with 374.33 kcal/100g, while DA1-DA5 with 352.67 – 371.0 kcal/100g, DVP (368.67 kcal/100g) and DE (366.0 kcal/100g), respectively.

Table 2 Proximate Composition of *Dioscorea* spp. collected in Peninsular Malaysia.

Sample	Ash	Fat	Protein	Carbohydrate	Energy
			g / 100 g		kcal / 100g
DA1	3.07 ^b	0.10 ^{cd}	7.43 ^c	82.23 ^d	358.67 ^f
DA2	2.23 ^d	0.13 ^c	10.57 ^a	80.17 ^e	364.0 ^e
DA3	2.73 ^c	0.00 ^d	8.0 ^b	84.67 ^b	371.0 ^b
DA4	2.73 ^c	0.40 ^b	7.23 ^c	80.03 ^e	352.67 ^g
DA5	2.87 ^c	0.53 ^a	6.0 ^e	83.60 ^c	363.33 ^e
DVP	1.90 ^e	0.17 ^c	8.03 ^b	83.77 ^c	368.67 ^c
DE	3.83 ^a	0.57 ^a	6.50 ^d	83.67 ^c	366.0 ^d
DP	1.60 ^f	0.00 ^d	2.77 ^f	90.80 ^a	374.33 ^a

Means followed by the same letter within a column are not significantly different at ($P \leq 0.05$) by Duncan's Multiple Range Test (DMRT) (Mean SEM, $n=3$). *, ** and *** significantly difference at $P < 0.05$, 0.01 and 0.001 respectively and NS= not significant.

The carbohydrate content in all accessions of *Dioscorea* spp. does not vary from the previously published data. Five different varieties of *D. alata* also showed a range of carbohydrate content ranging from 70.88 to 73.90 g/100g (Oko & Famurewa 2015), while research conducted by Ezeabara & Anona et al. (2018) showed that the carbohydrate content of *D. alata* in Nigeria was 76.26 g/100g. Meanwhile, the carbohydrate content in different varieties of *Dioscorea* spp. showed carbohydrate content ranging from 20.46 g/100g to 92.50 g/100g DW (Kulasinghe & Ranaweera, 2019). Additionally, recent research by Godfrey et al. (2023) on comparative assessment of the proximate composition in *D. alata*, *D. rotunda*, and *D. bulbifera* revealed the carbohydrate composition in the range of 57.38 to 77.51g/100g.

Meanwhile, DA2 showed the highest protein content (10.57 g/100g) ($p < 0.05$), while DP had the lowest protein content (2.77 g/ 100g). These findings and values agreed with the previous data on nutritional compositions of various landraces of *Dioscorea* spp. Twenty-five landraces of *D. alata*, *D. opposita*, *D. fordii* and *D. persimilis* in China were studied with a protein content range from 6.3% to 12.2%, while 36 yams landraces were studied in Southwest Ethiopia with protein content from 6.25% to 8.28% (Wu et al., 2016; Mulualem et al., 2018). A study by Padhan et al. (2020) on eight wild yams and one cultivated *Dioscorea* revealed a high protein content in the wild yam of *D. pubera* with 10.3%. Our findings were also aligned with the theory that different accessions of yams contribute to different nutritional compositions. For example, studies on five cultivars of *D. alata* flour fractions (yellow yam, orange

yam, light purple yam, purple yam and dark purple yam) were compared, and the protein content in various flour fractions revealed light purple of *D. alata* had the highest of protein content (9.85%)(Nadia et al. 2015).

Phytochemical Analysis

Meanwhile, the total phenolic content from the methanolic extract of various accessions of *Dioscorea* spp. is shown in Table 3. *D. piscatorum* (DP) had the highest content of phenolics (300.55 mg GAE/100g)

($p < 0.05$) compared to other accessions. Meanwhile, the phenolics content in all accessions of *D. alata* (light yellow variation, DA1-DA5) exhibited the amount of phenolics ranging from 38.9 mg GAE/100g - 74.1 mg GAE/100g, with DA3 showing the highest amount of phenolics content (74.1 mg GAE/100g). Meanwhile, the methanolic extracts of DVP and DE displayed phenolic content of 60.92 mg GAE/100g and 84.70 mg GAE/100g, respectively.

Table 3. Total Phenolic, Total Flavonoid, Flavonoid/Phenolic Ratio and Inhibition Concentration of *Dioscorea* spp.

Sample	Total Phenolic (mg GAE/100 g)	Total Flavonoid (mg QE/100 g)	Flavonoid/Phenolic Ratio	Inhibition Concentration (IC ₅₀ mg/ml)
DA1	38.90 ^d	11.97 ^d	0.31	7.31 ^b
DA2	39.52 ^d	19.51 ^d	0.49	15.61 ^h
DA3	74.10 ^b	38.53 ^c	0.52	9.55 ^d
DA4	49.85 ^{cd}	20.05 ^d	0.40	8.13 ^c
DA5	43.44 ^d	12.98 ^d	0.30	15.07 ^g
DVP	60.92 ^c	43.76 ^{bc}	0.72	9.69 ^e
DE	84.70 ^b	54.52 ^b	0.64	14.59 ^f
DP	300.55 ^a	183.03 ^a	0.61	4.73 ^a

Means followed by the same letter within a column are not significantly different at ($P \leq 0.05$) by Duncan's Multiple Range Test (DMRT) (Mean SEM, $n=3$). *, ** and *** significantly difference at $P < 0.05$, 0.01 and 0.001 respectively and NS= not significant. IC₅₀ = Inhibition concentration at which DPPH radicals were scavenged by 50%.

The methanolic extracts of DP, DE and DVP demonstrated the highest flavonoid content with 183.03 mg QE/100g, 54.52 mg QE/100g and 43.76 mg QE/100g, respectively. The flavonoid/phenolic ratio in the DVP was high compared to other accessions, with 0.72, followed by DE (0.64) and DP (0.61), while DA1-DA5 had values ranging from 0.30 to 0.52. The variations of phytochemicals in different accessions of *Dioscorea* spp. due to the many contributing factors. The dark purple hue of the flesh from DVP is due to the high amount of anthocyanin pigment, which is then proven to have the highest flavonoid/phenolic ratio across all the accessions. Alatanin, a class of cyanidin in anthocyanins, was identified as a significant chemical compound present in *D. alata* (purple variety) in Thailand and the Philippines (Moriya et al., 2015; Srivichai & Hongprabhas 2020). Meanwhile, the study of phenolic content from different landraces of *Dioscorea* spp. including *D. esculenta* (highland and lowland) in the Philippines was conducted, whereby the phenolics content in the highland variety showed an impeccable higher phenolics content as compared to the lowland with 156.5 mg GAE/100g and 112. 4 mg GAE/100g, respectively (Cornago et al., 2011). The development of the HP-TLC method to quantify the secondary metabolites in the *D. esculenta* in Vanuatu showed the presence of epicatechin, a flavonoid class of compound, along with saponin (Lebot et al., 2019). Additionally, the

effects of pigment colours from the different landraces of *D. alata* in India and their correlation with the phenolics content showed that the yellow and purple flesh tuber share higher phenolics content and their antioxidant activity (Jose et al., 2019).

Meanwhile, the in vitro antioxidant activity was measured using a free radical scavenging assay on eight extracts of *Dioscorea* spp, and a significant difference in the inhibition concentration was revealed. The methanolic extracts of DA1, DA3, DA4, DVP and DP showed IC₅₀ values < 10 mg/ml, with DP having the lowest IC₅₀ value of 4.73 mg/ml ($p < 0.05$), indicating a more potent antioxidant activity compared to the others accessions. However, the flavonoid/phenolic ratio in DP is lower compared to DVP. Still, it exhibits the most potent antioxidant activity (IC₅₀ = 4.73 mg/ml), probably due to the presence of carotenoids, which is attributed to its dark yellow-orangish hue of the flesh tuber that is present in various accessions of *Dioscorea* spp. (Nadia et al., 2015; Jose et al., 2019; Ukom et al., 2020).

CONCLUSIONS

The studies on different accessions of *Dioscorea* spp. showed different proximate compositions, phenolics and flavonoid content. The free radical scavenging assay towards methanolic extracts also showed diverse

antioxidant activities depending on their varieties and localities. *D. piscatorium* (DP) showed the highest carbohydrate content and provided more energy than other accessions. This accession exhibits exceptional antioxidant activity that could benefit health applications emphasizing oxidative stress reduction. Meanwhile, despite the high protein content in the variety of *D. alata* (DA2), its antioxidant activity was weak compared to other accessions. In conclusion, the samples vary significantly in both nutritional and functional properties. These differences highlight the suitability of each sample for specific dietary or health-related applications.

Acknowledgements: The author wishes to extend their deepest gratitude to the Herbarium (Kompleks MyGene Bank) for identifying the samples and the Alliance Chemist Laboratory Sdn. Bhd. for conducting the proximate analysis.

Authors' Contributions: All authors contributed to the manuscript. Mohd Zulkhairi Azid designed and assisted with the experiments, visualized data, and drafted, compiled, and edited the manuscript. Mohd Norfaizal Ghazalli and Salmaniza Salleh collected and identified all sample materials. Abdul Muhaimin Abdul Kadir assisted with data visualization. Siti Aisyah Mohd Noor, Saidatul Aqilah Mohamad Yusof, Nur Daliana Yusof, and Mohammad Raiehan Prayoga contributed to sample preparation and experimental design. All authors have read and approved the final manuscript.

Competing Interests: All authors declared that they have no competing interests in this study.

Funding: The research was made possible by funding from the Ministry of Agriculture and Food Security (KPKM), Malaysia, for the entire duration of the study.

REFERENCES

- Azahana, A., Dahlia, A.S., Noor Syaheera, M.Y., Noor Haslinda, N.H., Othman, H.S., & Salleh, M.S. (2023). Foliar morphological and micromorphological variation of *Dioscorea* species in Terengganu, Malaysia. *Revelation and Science*, 1(1), 29–38. doi: <https://doi.org/10.31436/revival.v1i1.379>
- Adoménienė, A., & Venskutonis, P. R. (2022). *Dioscorea* spp.: Comprehensive review of antioxidant properties and their relation to phytochemicals and health benefits. *Molecules*, 27(8). doi: <https://doi.org/10.3390/molecules27082530>
- Cornago, D. F., Rumbaoa, R. G. O., & Geronimo, I. M. (2011). Philippine yam (*Dioscorea* spp.) tubers phenolic content and antioxidant capacity. *Philippine Journal of Science*, 140(2), 145–152.
- DOA. (2023). Department of Agriculture Booklet Statistik Tanaman (Sub Sektor Tanaman Makanan) 2023 (2024, December 3). Retrieved from https://www.doa.gov.my/doa/resources/aktiviti_sumber/sumbe_r_awam/maklumat_pertanian/perangkaan_tanaman/booklet_statistik_tanaman_2023.pdf
- Ezeabara, C. A., & Anona, R. O. (2018). Comparative analyses of phytochemical and nutritional compositions of four species of *Dioscorea*. *Acta Scientific Nutritional Health*, 2(7), 90–94.
- FAO STAT. Food and Agriculture Organization of the United Nations (2024, November 11). Retrieved from <https://www.fao.org/faostat/en/#data/QCL>
- Ferriol, M. (2023). Morphological diversity of edible yam (*Dioscorea* spp) in Tablas, Island, Romblon, Philippines. *Romblon State University Research Journal*, 5(1), 33–42. doi: <https://doi.org/10.58780/rsurj.v5i1.89>
- Godfrey, E. O., Faith, O., & Esther, I. I. (2023). Comparative assessment of the proximate composition, functional properties and amino acid profile of *Dioscorea bulbifera*, *Dioscorea alata* and *Dioscorea rotundata* found in Minna, Niger State. *Biology, Medicine, & Natural Product Chemistry*, 12(1), 177–185. doi: <https://doi.org/10.14421/biomedich.2023.121.177-185>
- Induar, S., Dubey, D., Rath, S., Meher, R. K., Swain, S. K., & Tripathy, S. K. (2024). Evaluation of the antioxidant and antimicrobial activity of the nutritionally rich plant, *Dioscorea alata* L. *Biomedical and Pharmacology Journal*, 17(2), 1265–1278. doi: <https://doi.org/10.13005/bpj/2940>
- Islam, A., Chakraborty, D., Bhattacharjee, S. C., Ali, H., Sarker, F., & Islam, S. M. M. (2024). Assessment of total phenolic content (TPC), total flavonoid content (TFC) and antidiarrheal and antioxidant activities of *Dioscorea bulbifera* tuber extract. *Asian Journal of Pharmaceutical and Clinical Research*, 17(11), 195–199. doi: <https://doi.org/10.22159/ajpcr.2024v17i11.51957>
- Jose, A., Nizar, S., Vishnu, M. R., & Anil Kumar, M. (2019). Antioxidant potential in relation to total phenolics and pigment contents among the selected landraces of *Dioscorea alata* L. In Kerala, India. *Journal of Experimental Biology and Agricultural Sciences*, 7(4), 403–410. doi: [https://doi.org/10.18006/2019.7\(4\).403.410](https://doi.org/10.18006/2019.7(4).403.410)
- Kuagny, R. B. M., Saha, B. U. F., Achu, M. B. L., Kansci, G., Medoua, G. N., & Fokou, E. (2023). Polyphenol and flavonoid contents, antioxidant activities and glycaemic index of *Dioscorea* spp and *Coleus esculentus* from Cameroon. *Food Chemistry Advances*, 3(June), 100500. doi: <https://doi.org/10.1016/j.focha.2023.100500>
- Kulasinghe, W. M. A. A., & Ranaweera, K. K. T. N. (2019). Physical, chemical and biological aspects of *Dioscorea* yams and potential value additions. *Journal of Agriculture and Value Addition*, 2(1), 43–59. doi: <https://doi.org/10.4038/java.v2i1.109>
- Lebot, V., Malapa, R., & Molisálé, T. (2019). Development of HP-TLC method for rapid quantification of sugars, catechins, phenolic acids and saponins to assess yam (*Dioscorea* spp.) tuber flour quality. *Plant Genetic Resources: Characterisation and Utilisation*, 17(1), 62–72. doi: <https://doi.org/10.1017/S1479262118000333>
- Li, T., Teng, H., An, F., Huang, Q., Chen, L., & Song, H. (2019). The beneficial effects of purple yam (*Dioscorea alata* L.) resistant starch on hyperlipidemia in high-fat-fed hamsters. *Food and Function*, 10(5). doi: <https://doi.org/10.1039/c8fo02502a>
- Li, Y., Ji, S., Xu, T., Zhong, T., Xu, M., Liu, X., Li, M., Fan, B., Wang, F., Xiao, J., & Lu, B. (2023). Chinese yam (*Dioscorea*): Nutritional value, beneficial effects, and food and pharmaceutical application. *Trends in Food Science &*

- Technology, 134, 29-40.
doi:https://doi.org/10.1016/j.tifs.2023.01.021
- Logan, K., Nwokocha, C., Asemota, H., & Gray, W. (2024). Characterization of ACE inhibitory activity in *Dioscorea alata* cv and its implication as a natural antihypertensive extract. *Journal of Ethnopharmacology*, 319(2), 117221. doi:https://doi.org/10.1016/j.jep.2023.117221
- Mainasara, M. M., Abu Bakar, M. F., Md Akim, A., Linatoc, A. C., Abu Bakar, F. I., & Ranneh, Y. K. H. (2021). Secondary metabolites, antioxidant, and antiproliferative activities of *Dioscorea bulbifera* leaf collected from Endau Rompin, Johor, Malaysia. *Evidence-Based Complementary and Alternative Medicine*, 2021. doi:https://doi.org/10.1155/2021/8826986
- Mirfat, A. H. S., Razali, M., & Salma, I. (2013). Antioxidant properties of wild *Mangifera* species in Malaysia. *Acta Horticulturae*, 979, 651–660. doi:https://doi.org/10.17660/ActaHortic.2013.979.72
- Mojiono, M., Ajiid, M. D. S., Wulandari, S., Apriliyanti, F., Alamsyah, F., & Fansuri, H. (2022). Profile of yam tubers (*Dioscorea* spp.) and its potential in food products: A mini review. *Jurnal Agrosains : Karya Kreatif Dan Inovatif*, 7(1), 36–41. doi:https://doi.org/10.31102/agrosains.2022.7.1.36-41
- Moriya, C., Hosoya, T., Agawa, S., Sugiyama, Y., Kozono, I., Shin-Ya, K., Terahara, N., & Kumazawa, S. (2015). New acylated anthocyanins from purple yam and their antioxidant activity. *Bioscience, Biotechnology and Biochemistry*, 79(9), 1484–1492. doi:https://doi.org/10.1080/09168451.2015.1027652
- Muluallem, T., Mekbib, F., Hussein, S., & Gebre, E. (2018). Analysis of biochemical composition of yams (*Dioscorea* spp.) landraces from Southwest Ethiopia. *Agrotechnology*, 07(01). doi:https://doi.org/10.4172/2168-9881.1000177
- N'dri, G., Digbeu, Y. D., Makambou, J. G., & Dué, E. A. (2018). The impact of cooking on the proximate composition and antinutritional factors of yam *Dioscorea esculenta* tubers. *International Journal of Trend in Research and Development*, 5, 456–460.
- Nadia, L., Aman Wirakartakusumah, M., Andarwulan, N., Purnomo, E. H., Noda, T., & Ishiguro, K. (2015). Chemical characterization of flour fractions from five yam (*Dioscorea alata*) cultivars in Indonesia. *Journal of Engineering and Technological Sciences*, 47(1), 92–103. doi:https://doi.org/10.5614/j.eng.technol.sci.2015.47.1.7
- Nurul Makiyah, S. N., Christina, Y. I., & Ahkam, A. H. (2023). *Dioscorea alata* extract as anti-cancer agent by regulating genetic mutations, apoptosis, and cell proliferation: in vitro and in silico Studies. *Sains Malaysiana*, 52(9), 2657–2671. doi:https://doi.org/10.17576/jsm-2023-5209-15
- Oko, A., & Famurewa, A. (2015). Estimation of nutritional and starch characteristics of *Dioscorea alata* (water yam) varieties commonly cultivated in the South-Eastern Nigeria. *British Journal of Applied Science & Technology*, 6(2), 145–152. doi:https://doi.org/10.9734/bjast/2015/14095
- Padhan, B., Biswas, M., & Panda, D. (2020). Nutritional, antinutritional and physico-functional properties of wild edible yam (*Dioscorea* spp.) tubers from Koraput, India. *Food Bioscience*, 34, 100527. doi:https://doi.org/10.1016/j.fbio.2020.100527
- Povydysh, M. N., Titova, M. V., Ivkin, D. Y., Krasnova, M. V., Vasilevskaya, E. R., Fedulova, L. V., Ivanov, I. M., Klushin, A. G., Popova, E. V., & Nosov, A. M. (2023). The hypoglycemic and hypocholesterolemic activity of *Dioscorea deltoidea*, *Tribulus terrestris* and *Panax japonicus* cell culture biomass in rats with high-fat diet-induced obesity. *Nutrients*, 15(3). doi:https://doi.org/10.3390/nu15030656
- Price, E. J., Bhattacharjee, R., Lopez-Montes, A., & Fraser, P. D. (2018). Carotenoid profiling of yams: Clarity, comparisons and diversity. *Food Chemistry*, 259, 130–138. doi:https://doi.org/10.1016/j.foodchem.2018.03.066
- Rayamajhi, A., Karki, D., Phunyal, A., Sapkota, A., Adhikari, B., & Adhikari, A. (2024). Phytochemical profiling from *Dioscorea bulbifera* L. bulbils using LC-MS, proximate analysis and antidiabetic activity: in vitro and in silico approaches. *International Journal of Food Properties*, 27(1), 1396–1414. doi:https://doi.org/10.1080/10942912.2024.2410469
- Ren, Y., Chen, Y., Hu, B., Wu, H., Lai, F., & Li, X. (2015). Microwave-assisted extraction and a new determination method for total steroid saponins from *Dioscorea zingiberensis* C.H. Wright. *Steroids*, 104, 145–152. doi:https://doi.org/10.1016/j.steroids.2015.09.008
- Safitri, E., Humaira, H., Nazaruddin, N., Susilawati, S., Murniana, M., & Md Sani, N. D. (2021). *Dioscorea alata* L anthocyanin extract methanol as a sensitive pH active compound. *Journal of Physics: Conference Series*, 1869(1). doi:https://doi.org/10.1088/1742-6596/1869/1/012058
- Salahuddin, M. A. H., Ismail, A., Kassim, N. K., Hamid, M., & Ali, M. S. M. (2020). Phenolic profiling and evaluation of in vitro antioxidant, α -glucosidase and α -amylase inhibitory activities of *Lepisanthes fruticosa* (Roxb) Leenh fruit extracts. *Food Chemistry*, 331(April), 127240. doi:https://doi.org/10.1016/j.foodchem.2020.127240
- Saleha, S., Saidi, N., S., M., Rasnovi, S., & Iqbalsyah, T. M. (2018). Nutritional composition of *Dioscorea hispida* from different locations around Leuser ecosystem area. *Jurnal Natural*, 18(1), 1–6. doi:https://doi.org/10.24815/jn.v18i1.8504
- Sato, K., Umekage, K., & Kumamoto, M. (2024). Comparison of nutritional values for *Dioscorea esculenta* grown in a cultivated sub-tropical region and a plastic greenhouse in a cold region. *South African Journal of Botany*, 169, 1–5. doi:https://doi.org/10.1016/j.sajb.2024.04.013
- Srivichai, S., & Hongprabhas, P. (2020). Profiling anthocyanins in Thai purple yams (*Dioscorea alata* L.). *International Journal of Food Science*. doi:https://doi.org/10.1155/2020/1594291
- Syahputra, H., Masfria, M., Amana, M. R., & Rivai, M. (2024). Isolation and identification of diosgenin and flavonoids compound from ethanol extract of *Dioscorea hispida* Dennst. tuber. *Indonesian Journal of Pharmaceutical and Clinical Research*, 7(1), 7–15. doi:https://doi.org/10.32734/ijdp.v7i1.15994
- Ukom, A. O., Ojmelukwe, P. C., & Ezeama, C. F. (2020). Proximate composition and carotenoid profile of yam (*Dioscorea* spp.) and cocoyam (*Xanthosoma maffa*) Scotch root tubers from Nigeria. *American Journal of Food and Nutrition*, 4, 1–10.
- Wu, Z. G., Jiang, W., Nitin, M., Bao, X. Q., Chen, S. L., & Tao, Z. M. (2016). Characterizing diversity based on nutritional and bioactive compositions of yam germplasm (*Dioscorea* spp.) commonly cultivated in China. *Journal of Food and Drug Analysis*, 24(2), 367–375. doi:https://doi.org/10.1016/j.jfda.2015.12.003
- Yalindua, A., Manampiring, N., Waworuntu, F., & Yalindua, F. Y. (2021). Physico-chemical exploration of yam flour (*Dioscorea alata* L.) as a raw material for processed cookies. *Journal of Physics: Conference Series*, 1968(1). doi:https://doi.org/10.1088/1742-6596/1968/1/012004

Zulkhairi, A. M., Aisyah, M. N. S., Razali, M., Syafini, G. N., Umikalsum, M. B., Athirah, A. A., Daliana, M. Y. N., & Rosali, H. (2021). Antioxidants Capacity, Phenolic and Oxalate Content from Two Varieties of *Solanum melongena* at Different Maturity Stages. *Asian Journal of Applied Chemistry*

Research, 8 (October 2020), 54–63.63.
doi:<https://doi.org/10.9734/ajacr/2021/v8i430198>